

Histological image processing and analysis

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Introduction

Biological image processing and analysis algorithms were created to evaluate diverse aspects of skin pathology in histological specimens of both human subjects and mouse models of human disease. These included:

- * Measuring integrity in picrosirius-stained skin imaged with crosspolar microscopy to evaluate loss of collagen integrity. We measured degradation in collagen organisation (Figure 1, 2) and changes in bundle thickness in an ageing series (Figure 3), as well as in a model of diabetic skin.
- A technique to quantify collagen dynamics by assessing Herovici staining of old and young collagen fibres in skin from ageing and diabetic subjects (Figure 4).
- Quantification techniques to assess skin layer morphometrics (adipocyte hyperplasia and hypertrophy, and depth of each skin layer) to facilitate high-throughput analysis. (Figure 5, 6)
- Development of a technique to measure skin surface texture through the automated analysis of human skin impressions created from sun exposed and sun protected sites (Figure 7).
- Evaluation of changes in each of the three human skin layers in terms of morphology and structure using a variety of computational techniques.

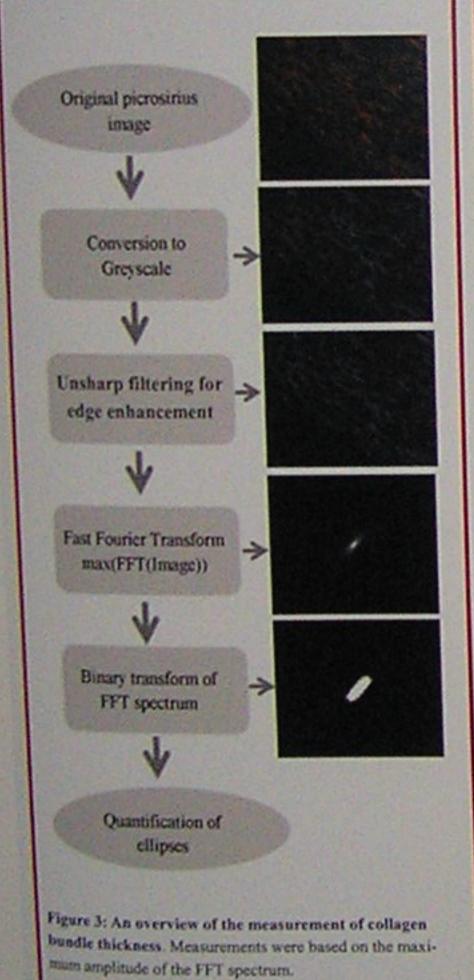


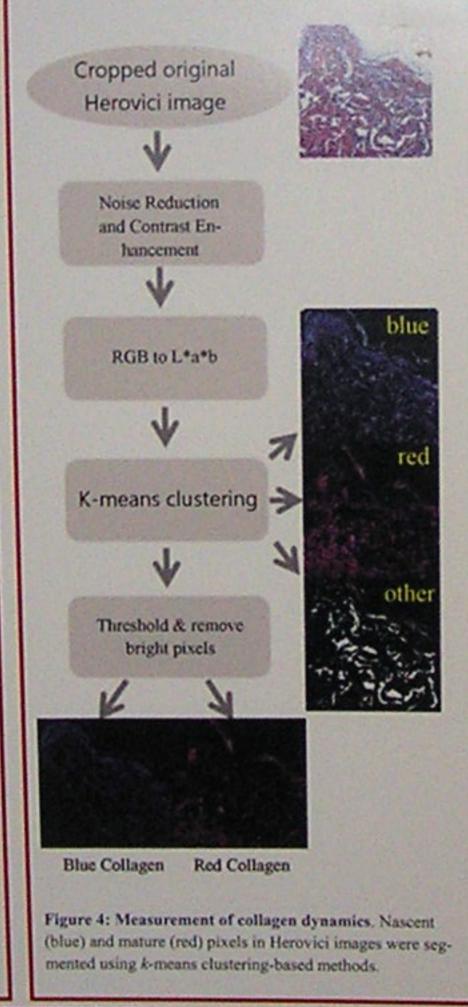
collagen images (left

owner sound and old skin.

a (exotte parallel and Bourier Transform (FFT)

Figure 2: Measuring collagen orientation in a picrosirius stained skin image (a). (b) FFT elliptical analysis: Gabor filtered image in different directions (left panels), FFT power spectra (centre panels) and ellipses generated from each direction (right panels).





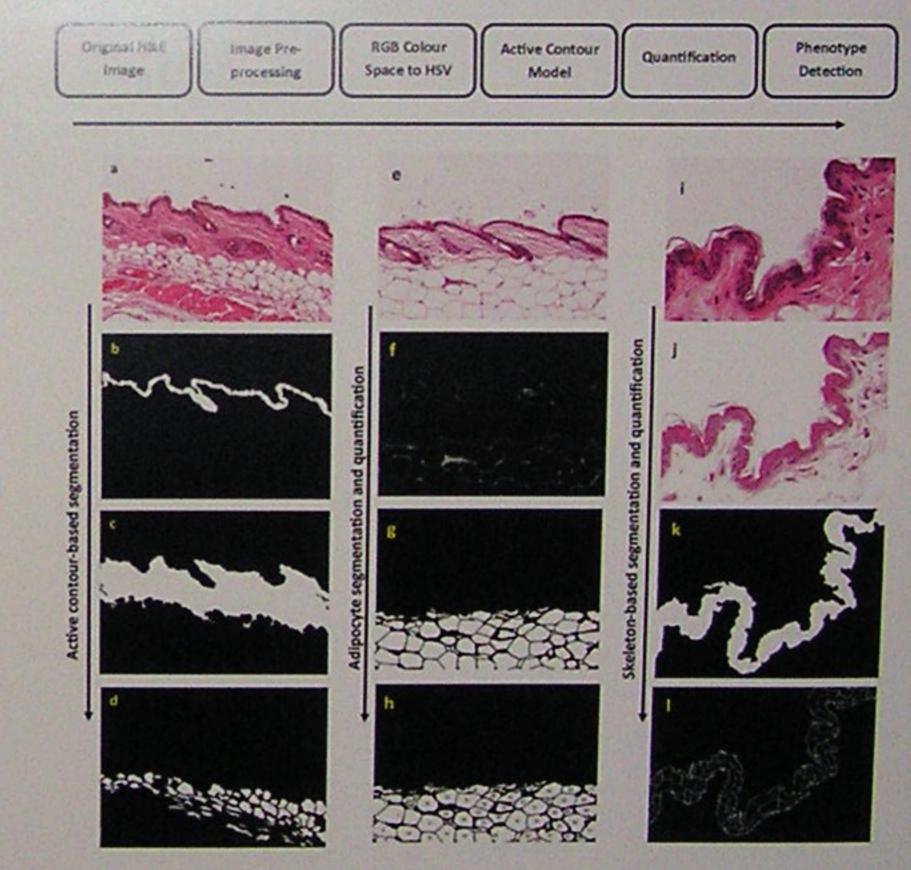
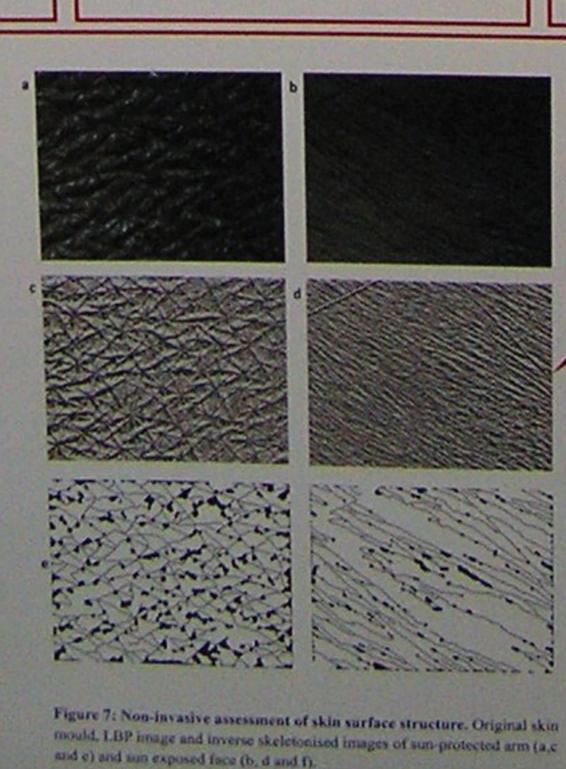


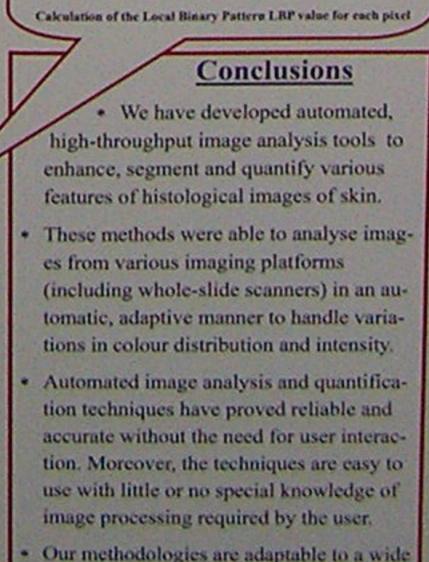
Figure 5: High-throughput cutaneous phenotype detection and quantification. Skin layer segmentation using an active contour method (a-d), quantification of adipocyte size and number (e-h), and epidermal and dermal layer thickness quantification (i-l).

Figure 6: Automated adipocyte size and number quantification. Adipocyte segmentation was achieved with piecewise fincar

transformation before counting and area

enconsupernerits.





without the need for user interaccover, the techniques are easy to
ittle or no special knowledge of

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Publications

Wargent CJ, Cawthorne MA, Jassim S, Langlands K.

A Novel Method to Assess Collagen Architecture in

Osman OS, Selway JL, Kępczyńska MA, Stocker CJ,

method for accurate adipocyte quantification. Adipo-

O'Dowd JF, Cawthorne MA, Arch JR, Jassim S,

Langlands K. A novel automated image analysis

cyte 2013; 2:160 - 164; http://dx.doi.org/10.4161/

* Osman OS, Selway JL, Harikumar PE, Jassim S and

Langlands K. Automated analysis of collagen histolo-

gy in ageing skin. In Proceedings of the International

Conference on Bioimaging, France, 3rd - 6th March

Skin. BMC Bioinformatics 2013

* Osman OS, Selway JL, Harikumar PE, Stocker CJ,

 Our methodologies are adaptable to a wide range of high-throughput morphometric projects.